Therapeutic Applications of Low-intensity Pulsed Ultrasound in Osteoporosis

Ali Yadollahpour¹, Samaneh Rashidi^{2*}

¹Department of Medical Physics, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran, ²Student Research Committee, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

Abstract

Ultrasound (US) waves due to their unique features can be a treatment option for osteoporosis. Initial studies have shown promising but controversial outcomes. Several preclinical and clinical studies have been conducted on osteoporosis and studies are ongoing to find optimum US parameters, mechanisms of action, and therapeutic efficacies of these techniques for osteoporosis treatment. This paper was aimed to review the recent advances of using US waves in the treatment of osteoporosis and possible mechanisms of actions. The databases of PubMed (1980-2016), EMBASE (1980-2016), Web of Sciences (1980-2016), and Google Scholar (1980-2016) were searched using the set terms. The obtained records were reviewed, and relevant studies were selected for comprehensive review of the current literature. Low-intensity pulsed US (LIPUS) has biological effects on the bone healing process and it can accelerate bone regeneration. Current evidence is limited on the efficacy of US waves for treatment or prevention of osteoporosis; however, the initial studies are promising. The US waves can promote osteoblast and inhibit osteoclast, enhance angiogenesis, trigger expression of different genes associated with osteogenesis. No definite dose-response existed on the clinical trials of US wave applications. The current evidence shows the therapeutic efficacy of US waves particularly LIPUS for osteoporosis treatment; however, to observe therapeutic outcomes long-term US stimulation is required. No definitive dose-response is proposed for osteoporosis. Further in vitro and clinical trials should be conducted to develop US-based techniques for the treatment of osteoporosis as a clinical treatment option.

Key words: Mechanism of action, osteoporosis, treatment, ultrasound

INTRODUCTION

steoporosis is among the common musculoskeletal disorders worldwide. The first choice of treatment for osteoporosis is currently medication. Bisphosphonates are the most common medications administered for osteoporosis treatment and prevention. The main types of these medications include alendronate (Fosamax), risedronate (Actonel), ibandronate (Boniva), and zoledronic acid (Reclast).^[1] Hormones, such as estrogen, and some hormone-like medications such as raloxifene (Evista) are also approved medications for preventing and treating The medications osteoporosis. have different side effects considering the relatively long-term administration of such medications.^[2] Moreover, fewer women use estrogen replacement therapy now because it may increase the risk of heart attacks and some types of cancer. Therefore, developing a new non-medication treatment for osteoporosis is necessary.

During the recent years, several non-medication treatments have been developed for the treatment of bone related disorders.^[3,4] At present, ultrasound (US) waves are known as therapeutic tools which are widely used in various fields of diagnostic and therapeutic medicine including structural and functional imaging, soft tissue injuries repairing, recovery of musculoskeletal anomalies and injuries, and reducing the pain.^[5-7] US is a mechanical longitudinal energy in the form of waves that can transfers mechanical energy into the tissues as a propagating pressure wave.^[8,9] Bone regeneration

Address for correspondence:

Department of Medical Physics, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran. E-mail: samanehrashidi92@gmail.com

Received: 06-10-2016 **Revised:** 09-01-2017 **Accepted:** 17-01-2017 involves a complex process such as inflammation, cellular proliferation and differentiation, chemotaxis, synthesis of an extracellular matrix and finally remodeling.^[10,11] Although the cellular mechanisms of the US effects are not clearly understood, several in vitro and in vivo studies have demonstrated that low-intensity pulsed US (LIPUS) has biological effects on the bone-healing process and it can accelerate bone regeneration.^[12-15] The results of several histological studies have shown that LIPUS has an important influence on key functional activities of all major cell types involved in bone healing, such as osteoblasts, osteoclasts, chondrocytes, and mesenchymal stem cells.^[16,17] In addition, several studies showed a positive effect of US to increase the levels of intracellular calcium incorporation in cultures of differentiating bone and cartilage cells. Increases in calcium incorporation were modulated transforming growth factor (TGF)-beta and adenylate cyclase activity. LIPUS also has been shown to increase the intracellular calcium concentration in chondrocytes and increase the percentage of calcified cartilage. Others have suggested that LIPUS have a stimulatory effect on endochondral ossification. Concurrently, stimulation of endochondral ossification is due to stimulation of bone cell differentiation and calcified matrix production.^[18-21] In addition, it has been shown in an animal model that LIPUS leads to stimulation of vascular endothelial growth factor. Others have shown that LIPUS treatment increases the degree of vascularity, indication that US increases blood flow.^[22,23] This study reviewed the mechanism of US stimulation in osteoporosis treatment.

LIPUS AND OSTEOPOROSIS

Osteoporosis is the most common bone disorder and a major and growing health problem worldwide. Several risk factors are involved in the occurrence and progression of osteoporosis including aging, sedentary lifestyle and estrogen deficiency due to menopause, ovariectomy, and hormonal therapy.^[24-27]

Various medications such as estrogen, bisphosphonates, calcitonin, calcium, and vitamin D have been used in the treatment of osteoporosis for many years. Although they are mainly used therapeutically as bone resorption inhibitors, they have no significant long-term effects.

Numerous *in vivo* animal and clinical trials studies have shown that LIPUS because of its properties and positive effects on the generation and activation of bone cells is capable of accelerating and augmenting the healing of osteoporosis. LIPUS produces the pressure waves, which in duce biochemical and molecular events at the cellular level and whereby accelerated healing of osteoporosis.^[22,28-32] The results of biomechanical and histologic investigations prove that LIPUS have effects on bone mineral density and mechanical strength. They also concluded that LIPUS stimulates bone formation in distraction osteogenesis and acceleration of healing or strength.^[33,34] Various studies have shown that LIPUS not only prevent bone loss but also restore bone mass. The finding suggested that LIPUS therapy, if scaled for whole body use, has clear clinical benefits for the treatment of osteoporosis.^[22,35-37]

LIPUS IN CELLULAR LEVELS

Most of the experimental studies showed that LIPUS treatment influence cell membrane permeability and increase cellular activity.^[31,38-40]

Several genes such as alkaline phosphatase (ALP), bone sialoprotein (BSP), collagen type I, osteocalcin (OC), and osteonectin (ON) are characteristic of osteoblast differentiation. They are overexpressed during the process of osteogenesis.^[41] The results of a study that evaluated the genetic expression and response to LIPUS in rat osteoblastic cells showed early response genes in bone marrow-derived stromal cells. In this study, the gene expression level in LIPUS group is demonstrated and calculated over the control group. The results between LIPUS stimulated group and sham control group for cyclooxygenase-2, early growth response-1, TGF-beta stimulated clone-22, ON, and osteopontin had shown a statistically significant difference.^[42] Another study evaluated the effect of LIPUS on the differentiation of pluripotent mesenchymal cell line C2C12 by examining particular mRNA and protein expression levels. C2C12 cells have the capacity to differentiate into myoblasts, osteoblasts, chondroblasts, or adipocytes. The results determined that LIPUS stimulation increased Runx2 protein expression and phosphorylation of ERK1/2 and p38 mitogen-activated protein kinase (MAPK). They also demonstrated that LIPUS stimulation converts the differentiation pathway of C2C12 cells into the osteoblast and/or chondroblast lineage via activated phosphorylation of ERK1/2 and p38 MAPK.^[43] Mukai et al. (2005) in their in vitro study demonstrated that LIPUS promoted the mRNA expression of type II collagen, type X collagen, aggrecan, and TGF- β in rat chondrocytes.^[44] In addition, Chen et al. (2003) reported that LIPUS stimulation elevated Runx2 mRNA expression and gradually promoted OC mRNA expression in human osteoblasts.^[40] Other several in vitro studies had shown LIPUS elevated mRNA levels for insulin-like growth factor-I, OC, and BSP and also it was found to stimulate mRNA expression of the bone matrix proteins ALP and OC in UMR-106 cells.^[16,17] In addition, there have been some reports that LIPUS may have a direct effect on cell membrane permeability.[23,45-50]

TEMPERATURE VARIATIONS

The range of energies used in LIPUS treatment is relatively low which is the range of non-thermogenic and nondestructive. High-intensity US waves that are used in therapeutic and surgical applications (1-300 W/cm²) generates considerable heat in living tissue. Some of the investigators reported that the therapeutic benefits observed with LIPUS stimulation involve non-thermal mechanisms. Contrary, some researchers believe that the ability of LIPUS to stimulate changes in tissues and cells may be related to the temperature rising effects induced by energy absorption.^[51-54] High intensities (1000-3000 mW/cm²) have temperature effects and can cause considerable heating of tissues. Whereas the heating effect from LIPUS (20 to 50 mW/cm²) is estimated <11°C,^[54] some enzymes such as matrix metalloproteinase or interstitial collagenase have been shown to be very sensitive to small variations in temperature. Therefore, minimal heating effects may affect on them.[55,56] Chang et al. (2002) investigated thermal effects of US stimulation on fracture healing. They reported that difference between the microwave hyperthermia treated limbs and the sham-treated limb was not quite statistically significant. They have suggested that LIPUS stimulation could increase the new bone formation but its effects probably are not mediated via hyperthermia.^[54]

Other investigators suggested that the therapeutic benefits observed in tissues and cells after LIPUS stimulation may also be associated with nonthermal processes such as acoustic streaming and cavitation. They have suggested that cavitation mechanisms may due to an increase in protein and collagen synthesis observed in human fibroblasts after US stimulation.^[57-62]

EFFECTS OF VARIOUS INTENSITIES

To finding an optimal LIPUS protocol studies have examined several different low intensities and frequencies. Several studies have investigated the role of intensity in therapeutic effects and an attempt to determine the optimal LIPUS setting. High energy and intensity US through absorbed by tissues lead to increase tissues temperature and kill malignant cells.^[7]

To reduce pain and muscle spasms, to decrease joint stiffness, and to improve muscle mobility use applications of US in intensities of 1-3 W/cm^{2} .^[63]

The results of a study showed that the response of cells to the US is highly dependent on the intensity. With increasing US intensity to 120, 390, and 1490 mW/cm² expression of ALP showed progressively increased.^[64]

In a study, two different LIPUS intensities compared directly to investigate the relationship between intensity of and restoring the mechanical properties of a rat femora following fracture (50 and 100 mW/cm²). The results showed that the group was treated with 50 mW/cm² LIPUS intensity had significantly greater maximum torque and torsion stiffness compared to 100 mW/cm² treated femora and untreated controls.^[13]

Another parameter of the US signal that has also been investigated in stimulating bone osteogenesis is the frequency.

Several studies showed that the response of cells to the US is also dependent on the frequency. In a study therapeutic results of two protocols with different frequencies (1.5 MHz, 3.0 MHz) and constant intensity (500 mW/cm²) on rat fibulae fractures healing were compared. Results showed that protocol with 1.5 MHz frequency had more advanced radiographic and histological healing.[46] Results of another study demonstrated that there was no significant difference in therapeutic results of two protocols with different frequencies (1.5 MHz, 0.5 MHz) and constant intensity (30 mW/cm²). In this study maximum torque and torsional stiffness was investigated and both frequencies almost equally led to increase these parameters compared to untreated controls.[65] Another two protocols with different frequencies (1.5 MHz, 3.0 MHz) and constant intensity (500 mW/cm²) were investigated by Tsai et al. (1992). Results of the group that had been treated with 1.5 MHz showed significantly greater mineral apposition rates.^[66]

Based on the results obtained to stimulate osteogenesis experimentally and clinically frequency of 1.5 MHz has been more commonly used.

TIME-DEPENDENT EFFECT

There have been some reports that LIPUS shows positive effects on the healing of fresh fracture, nonunion and delayed union and also it accelerates bone maturation in distraction osteogenesis in clinical treatment and animals models.^[67-72]

On the other hand, there are some controversial results in regard to LIPUS is most effective during the lengthening phase, but the optimal timing of LIPUS has not been established. The researchers sought to determine the stage of fracture repair process that US have the greatest influence effects.

Several *in vivo* experimental studies investigated the effect of LIPUS stimulation on the various phase of fracture healing. Results of some studies suggested that LIPUS does not affect the remodeling phase of fracture healing. These results determined that LIPUS have a significant effect on the earlier inflammatory or accelerate to callus formation phases of healing.^[22,31,73-75]

In a study with a rat femoral fracture model, hard callus area, bone mineral content, mechanical torsion properties were measured at 4 different periods (1-8 days, 9-16 days, 17-24 days, 1-24 days) after expose to LIPUS, along with histologic analysis. The findings reported statistically significant increases in all measured parameters in all groups when compared with the control group. They suggested that LIPUS acts on some cellular reactions at each stage of the fracture healing process.^[73]

Azuma *et al.* (2001) in an experimental study measured mechanical and histological changes at different time periods during the healing process. They investigated the timing or duration of stimulation effect after 8 and 25 days of LIPUS treatment. They reported that the results had not shown significant effect associated with the timing or duration on the bone mineral content, but they also reported a significant increase in bone stiffness and maximum torque in LIPUS group.^[73]

CONCLUSION

Most of the studies showed a positive effect of US on bone healing. Numerous studies have proved the effect of LIPUS on bone regeneration, changes in bone mineral content and density. They have also demonstrated LIPUS increase callus formation, and its biological changes. Based on the results, investigators suggested that LIPUS therapy with smaller and continuous mechanical stress is more useful in preventing bone loss and bone remodeling in a clinical setting.^[13,38,40,54,64-66] LIPUS is affected without pain, without the need for hospitalization and it is portable by the patients. Between the methods available to enhance musculoskeletal disorders healing, US has suggested as a safe, practical, and effective treatment.^[22,28,34,35] In our opinion, extensive clinical and experimental and long-term studies investigating biophysical mechanisms of LIPUS are required.

REFERENCES

- Cummings SR, Kelsey JL, Nevitt MC, O'Dowd KJ. Epidemiology of osteoporosis and osteoporotic fractures. Epidemiol Rev 1985;7:178-208.
- Health UD, Services H. Bone Health and Osteoporosis: A Report of the Surgeon General. Rockville, MD: US Department of Health and Human Services, Office of the Surgeon General; 2004. p. 87.
- Yadollahpour A, Rashidi S. Therapeutic applications of electromagnetic fields in musculoskeletal disorders: A review of current techniques and mechanisms of action. Biomed Pharmacol J 2014;7:23-32.
- Yadollahpour A, Rashidi S. A review of electromagnetic field based treatments for different bone fractures. Biosci Biotechnol Res Asia 2014;11:611-20.
- Mostafa J, Ali Y, Zohre R, Samaneh R. Electromagnetic fields and ultrasound waves in wound treatment: A comparative review of therapeutic outcomes. Biosci Biotechnol Res Asia 2015;12:185-95.
- Yadollahpour A, Mostafa J, Samaneh R, Zohreh R. Ultrasound therapy for wound healing: A review of current techniques and mechanisms of action. J Pure Appl Microbiol 2014;8:4071-85.
- Burdette E, Svensson G, Lu XQ, Hansen J, Slayton M, Foard W, *et al.*, editors. Ultrasound Hyperthermia System for Breast Cancer Treatment. 1994 Proceedings

of IEEE Ultrasonics Symposium; 1994.

- Ziskin MC. Ultrasound. Applications of Ultrasound in Medicine—Comparison with other Modalities. New York: Springer; 1987. p. 49-59.
- Erdogan O, Esen E, Ustün Y, Kürkçü M, Akova T, Gönlüsen G, *et al.* Effects of low-intensity pulsed ultrasound on healing of mandibular fractures: An experimental study in rabbits. J Oral Maxillofac Surg 2006;64:180-8.
- 10. Harwood PJ, Ferguson DO. (ii) An update on fracture healing and non-union. Orthop Trauma 2015;29:228-42.
- 11. Bolander ME. Regulation of fracture repair by growth factors. Proc Soc Exp Biol Med 1992;200:165-70.
- Li JK, Chang WH, Lin JC, Ruaan RC, Liu HC, Sun JS. Cytokine release from osteoblasts in response to ultrasound stimulation. Biomaterials 2003;24:2379-85.
- Yang KH, Parvizi J, Wang SJ, Lewallen DG, Kinnick RR, Greenleaf JF, *et al.* Exposure to low-intensity ultrasound increases aggrecan gene expression in a rat femur fracture model. J Orthop Res 1996;14:802-9.
- Hantes ME, Mavrodontidis AN, Zalavras CG, Karantanas AH, Karachalios T, Malizos KN. Lowintensity transosseous ultrasound accelerates osteotomy healing in a sheep fracture model. J Bone Joint Surg Am 2004;86-A:2275-82.
- 15. Busse JW, Bhandari M, Kulkarni AV, Tunks E. The effect of low-intensity pulsed ultrasound therapy on time to fracture healing: A meta-analysis. CMAJ 2002;166:437-41.
- Naruse K, Mikuni-Takagaki Y, Azuma Y, Ito M, Oota T, Kameyama K, *et al.* Anabolic response of mouse bonemarrow-derived stromal cell clone ST2 cells to lowintensity pulsed ultrasound. Biochem Biophys Res Commun 2000;268:216-20.
- Warden SJ, Favaloro JM, Bennell KL, McMeeken JM, Ng KW, Zajac JD, *et al.* Low-intensity pulsed ultrasound stimulates a bone-forming response in UMR-106 cells. Biochem Biophys Res Commun 2001;286:443-50.
- Korstjens CM, Nolte PA, Burger EH, Albers GH, Semeins CM, Aartman IH, *et al.* Stimulation of bone cell differentiation by low-intensity ultrasound - A histomorphometric *in vitro* study. J Orthop Res 2004;22:495-500.
- Parvizi J, Parpura V, Kinnick R, Greenleaf J, Bolander M. Low intensity ultrasound increases intracellular concentration of calcium in chondrocytes. J Bone Joint Surg Br 1997;79:452.
- Ryaby J, Bachner E, Bendo J, Dalton P, Tannenbaum S, Pilla A. Low intensity pulsed ultrasound increases calcium incorporation in both differentiating cartilage and bone cell cultures. Trans Orthop Res Soc 1989;14:15.
- Ryaby J, Mathew J, Duarte-Alves P. Low intensity pulsed ultrasound affects adenylate cyclase and TGF-β synthesis in osteoblastic cells. Trans Orthop Res Soc 1992;17:590.
- 22. Leung KS, Lee WS, Tsui HF, Liu PP, Cheung WH. Complex tibial fracture outcomes following treatment

with low-intensity pulsed ultrasound. Ultrasound Med Biol 2004;30:389-95.

- Rawool NM, Goldberg BB, Forsberg F, Winder AA, Hume E. Power doppler assessment of vascular changes during fracture treatment with low-intensity ultrasound. J Ultrasound Med 2003;22:145-53.
- Tella SH, Gallagher JC. Prevention and treatment of postmenopausal osteoporosis. J Steroid Biochem Mol Biol 2014;142:155-70.
- 25. Pouilles JM, Tremollieres F, Ribot C. Effect of menopause on femoral and vertebral bone loss. J Bone Miner Res 1995;10:1531-6.
- 26. Melton LJ, Chrischilles EA, Cooper C, Lane AW, Riggs BL. How many women have osteoporosis? J Bone Miner Res 2005;20:886-92.
- D'Amelio P, Grimaldi A, Di Bella S, Brianza SZ, Cristofaro MA, Tamone C, *et al.* Estrogen deficiency increases osteoclastogenesis up-regulating T cells activity: A key mechanism in osteoporosis. Bone 2008;43:92-100.
- Siska PA, Gruen GS, Pape HC. External adjuncts to enhance fracture healing: What is the role of ultrasound? Injury 2008;39:1095-105.
- 29. Schortinghuis J, Bronckers AL, Stegenga B, Raghoebar GM, de Bont LG. Ultrasound to stimulate early bone formation in a distraction gap: A double blind randomised clinical pilot trial in the edentulous mandible. Arch Oral Biol 2005;50:411-20.
- El-Bialy TH, Elgazzar RF, Megahed EE, Royston TJ. Effects of ultrasound modes on mandibular osteodistraction. J Dent Res 2008;87:953-7.
- Leung KS, Cheung WH, Zhang C, Lee KM, Lo HK. Low intensity pulsed ultrasound stimulates osteogenic activity of human periosteal cells. Clin Orthop Relat Res 2004;253-9.
- Pilla AA, Mont MA, Nasser PR, Khan SA, Figueiredo M, Kaufman JJ, *et al.* Non-invasive low-intensity pulsed ultrasound accelerates bone healing in the rabbit. J Orthop Trauma 1990;4:246-53.
- Sakurakichi K, Tsuchiya H, Uehara K, Yamashiro T, Tomita K, Azuma Y. Effects of timing of low-intensity pulsed ultrasound on distraction osteogenesis. J Orthop Res 2004;22:395-403.
- Wu S, Kawahara Y, Manabe T, Ogawa K, Matsumoto M, Sasaki A, *et al.* Low-intensity pulsed ultrasound accelerates osteoblast differentiation and promotes bone formation in an osteoporosis rat model. Pathobiology 2009;76:99-107.
- 35. Guerino MR, Santi FP, Silveira RF, Luciano E. Influence of ultrasound and physical activity on bone healing. Ultrasound Med Biol 2008;34:1408-13.
- van der Windt DA, van der Heijden GJ, van den Berg SG, ter Riet G, de Winter AF, Bouter LM. Ultrasound therapy for musculoskeletal disorders: A systematic review. Pain 1999;81:257-71.
- 37. Claes L, Rüter A, Mayr E. Low-intensity ultrasound enhances maturation of callus after segmental transport.

Clin Orthop Relat Res 2005;189-94.

- Yang RS, Lin WL, Chen YZ, Tang CH, Huang TH, Lu BY, *et al.* Regulation by ultrasound treatment on the integrin expression and differentiation of osteoblasts. Bone 2005;36:276-83.
- 39. Ebisawa K, Hata K, Okada K, Kimata K, Ueda M, Torii S, *et al.* Ultrasound enhances transforming growth factor beta-mediated chondrocyte differentiation of human mesenchymal stem cells. Tissue Eng 2004;10:921-9.
- 40. Chen YJ, Wang CJ, Yang KD, Chang PR, Huang HC, Huang YT, *et al.* Pertussis toxin-sensitive Galphai protein and ERK-dependent pathways mediate ultrasound promotion of osteogenic transcription in human osteoblasts. FEBS Lett 2003;554:154-8.
- 41. Stein GS, Lian JB, Stein JL, Van Wijnen AJ, Montecino M. Transcriptional control of osteoblast growth and differentiation. Physiol Rev 1996;76:593-629.
- 42. Sena K, Leven RM, Mazhar K, Sumner DR, Virdi AS. Early gene response to low-intensity pulsed ultrasound in rat osteoblastic cells. Ultrasound Med Biol 2005;31:703-8.
- 43. Ikeda K, Takayama T, Suzuki N, Shimada K, Otsuka K, Ito K. Effects of low-intensity pulsed ultrasound on the differentiation of C2C12 cells. Life Sci 2006;79:1936-43.
- Mukai S, Ito H, Nakagawa Y, Akiyama H, Miyamoto M, Nakamura T. Transforming growth factor-beta1 mediates the effects of low-intensity pulsed ultrasound in chondrocytes. Ultrasound Med Biol 2005;31:1713-21.
- 45. Padilla F, Puts R, Vico L, Raum K. Stimulation of bone repair with ultrasound: A review of the possible mechanic effects. Ultrasonics 2014;54:1125-45.
- 46. Dyson M, Brookes M. Stimulation of bone repair by ultrasound. Ultrasound Med Biol 1983;Suppl 2:61-6.
- 47. Mortimer AJ, Dyson M. The effect of therapeutic ultrasound on calcium uptake in fibroblasts. Ultrasound Med Biol 1988;14:499-506.
- 48. Dinno MA, Dyson M, Young SR, Mortimer AJ, Hart J, Crum LA. The significance of membrane changes in the safe and effective use of therapeutic and diagnostic ultrasound. Phys Med Biol 1989;34:1543-52.
- 49. Ryaby J. Low intensity pulsed ultrasound modulates adenylate cyclase activity and transforming growth factor beta synthesis. Electomagnetics in Biology and Medicine. San Francisco: San Francisco Press; 1991.
- 50. Claes L, Willie B. The enhancement of bone regeneration by ultrasound. Prog Biophys Mol Biol 2007;93:384-98.
- 51. Wu JR, Du GH. Temperature elevation generated by a focused Gaussian ultrasonic beam at a tissue-bone interface. J Acoust Soc Am 1990;87:2748-55.
- Moros EG, Novak P, Straube WL, Kolluri P, Yablonskiy DA, Myerson RJ. Thermal contribution of compact bone to intervening tissue-like media exposed to planar ultrasound. Phys Med Biol 2004;49:869-86.
- 53. Fujii M, Sakamoto K, Toda Y, Negishi A, Kanai H. Study of the cause of the temperature rise at the muscle-bone interface during ultrasound hyperthermia. IEEE Trans Biomed Eng 1999;46:494-504.

- Chang WH, Sun JS, Chang SP, Lin JC. Study of thermal effects of ultrasound stimulation on fracture healing. Bioelectromagnetics 2002;23:256-63.
- 55. Welgus HG, Jeffrey JJ, Eisen AZ. The collagen substrate specificity of human skin fibroblast collagenase. J Biol Chem 1981;256:9511-5.
- 56. Welgus HG, Jeffrey JJ, Eisen AZ, Roswit WT, Stricklin GP. Human skin fibroblast collagenase: Interaction with substrate and inhibitor. Coll Relat Res 1985;5:167-79.
- 57. Hill C. Ultrasonic exposure thresholds for changes in cells and tissues. J Acoust Soc Am 1972;52:667-72.
- Dyson M. Non-thermal cellular effects of ultrasound. Br J Cancer Suppl 1982;5:165-71.
- 59. Feril LB Jr, Kondo T, Zhao QL, Ogawa R. Enhancement of hyperthermia-induced apoptosis by non-thermal effects of ultrasound. Cancer Lett 2002;178:63-70.
- Harvey W, Dyson M, Pond JB, Grahame R. The stimulation of protein synthesis in human fibroblasts by therapeutic ultrasound. Rheumatol Rehabil 1975;14:237.
- 61. Webster DF, Harvey W, Dyson M, Pond JB. The role of ultrasound-induced cavitation in the *in vitro* stimulation of collagen synthesis in human fibroblasts. Ultrasonics 1980;18:33-7.
- 62. Brujan EA. The role of cavitation microjets in the therapeutic applications of ultrasound. Ultrasound Med Biol 2004;30:381-7.
- 63. ter Haar G. Therapeutic applications of ultrasound. Prog Biophys Mol Biol 2007;93:111-29.
- Harle J, Salih V, Knowles JC, Mayia F, Olsen I. Effects of therapeutic ultrasound on osteoblast gene expression. J Mater Sci Mater Med 2001;12:1001-4.
- 65. Wang SJ, Lewallen DG, Bolander ME, Chao EY, Ilstrup DM, Greenleaf JF. Low intensity ultrasound treatment increases strength in a rat femoral fracture model. J Orthop Res 1994;12:40-7.
- 66. Tsai CL, Chang WH, Liu TK. Preliminary studies of duration and intensity of ultrasonic treatments on fracture repair. Chin J Physiol 1992;35:21-6.

- Heckman JD, Ryaby JP, McCabe J, Frey JJ, Kilcoyne RF. Acceleration of tibial fracture-healing by non-invasive, low-intensity pulsed ultrasound. J Bone Joint Surg Am 1994;76:26-34.
- Kristiansen TK, Ryaby JP, McCabe J, Frey JJ, Roe LR. Accelerated healing of distal radial fractures with the use of specific, low-intensity ultrasound. A multicenter, prospective, randomized, double-blind, placebocontrolled study. J Bone Joint Surg Am 1997;79:961-73.
- 69. Mayr E, Frankel V, Rüter A. Ultrasound An alternative healing method for nonunions? Arch Orthop Trauma Surg 2000;120:1-8.
- Takikawa S, Matsui N, Kokubu T, Tsunoda M, Fujioka H, Mizuno K, *et al.* Low-intensity pulsed ultrasound initiates bone healing in rat nonunion fracture model. J Ultrasound Med 2001;20:197-205.
- Shimazaki A, Inui K, Azuma Y, Nishimura N, Yamano Y. Low-intensity pulsed ultrasound accelerates bone maturation in distraction osteogenesis in rabbits. J Bone Joint Surg Br 2000;82:1077-82.
- 72. Mayr E, Laule A, Suger G, Rüter A, Claes L. Radiographic results of callus distraction aided by pulsed low-intensity ultrasound. J Orthop Trauma 2001;15:407-14.
- 73. Azuma Y, Ito M, Harada Y, Takagi H, Ohta T, Jingushi S. Low-intensity pulsed ultrasound accelerates rat femoral fracture healing by acting on the various cellular reactions in the fracture callus. J Bone Miner Res 2001;16:671-80.
- 74. Gebauer D, Mayr E, Orthner E, Ryaby JP. Low-intensity pulsed ultrasound: Effects on nonunions. Ultrasound Med Biol 2005;31:1391-402.
- 75. Rubin C, Bolander M, Ryaby JP, Hadjiargyrou M. The use of low-intensity ultrasound to accelerate the healing of fractures. J Bone Joint Surg Am 2001;83-A:259-70.

Source of Support: This study was financially supported by Student Research Committee, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran (No.:94S65). **Conflict of Interest:** Nil.